

The Correct Structure of Aquatolide—Experimental Validation of a Theoretically-Predicted Structural Revision

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Supporting Information

ABSTRACT: Aquatolide has been reisolated from its natural source, and its structure has been revised on the basis of quantum-chemical NMR calculations, extensive experimental NMR analysis, and crystallography.

A quatolide is a humulane-derived sesquiterpenoid lactone isolated from *Asteriscus aquaticus*.¹ The structure of aquatolide originally proposed on the basis of 1D and 2D NMR analysis (**1a**, Figure 1) contains an exceedingly rare

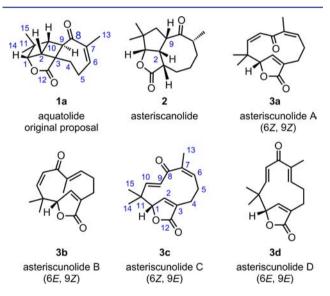


Figure 1. Originally proposed structure of aquatolide and structures of the related natural products asteriscanolide and asteriscunolides A–D.

[2]ladderane substructure.^{1–3} Intrigued by this structural unit, we initiated quantum-chemical calculations to verify the reported connectivity in preparation for studies of the biogenesis of aquatolide in nature.^{4–6} As described below, however, these seemingly innocuous calculations set us on a journey toward an extensive structural revision of this complex natural product.

Figure 1 also shows the structures of the related natural products asteriscanolide $(2)^7$ and asteriscunolides $A-D^8$ (3a-d, putative precursors to aquatolide via a photochemical 2 + 2 reaction), all isolated from the same plant. 3a-d have since been

found to display anticancer activity against several cell lines, which in part motivated the recent total synthesis of the D isomer.⁹ **2** has received much attention from the synthetic organic community, mainly because of its unusual carbon skeleton, and several successful syntheses have been reported in the literature.¹⁰ Studies of the synthesis or biosynthesis of aquatolide have not been described in the literature.

As shown in Table 1, many of the computed chemical shifts for structure 1a did not match up well with those reported for the natural product. Significant deviations were found for nuclei throughout the molecule, indicating that the core structure was likely incorrect. It is also noteworthy that experimentally, all three methyl carbon chemical shifts lie within a 1 ppm range, while the computed values for the methyl carbons in the proposed structure span an 8.5 ppm range. While the proposed structure was consistent with all of the reported experimental data, including nuclear Overhauser effect (NOE) and other correlations, our high-accuracy chemical shift calculations indicated that the assigned structure was incorrect.

The problem of identifying the correct structure of aquatolide thus arose. Our efforts consisted of a combination of both rational and arbitrary changes to the structure followed by recalculation of the ¹H and ¹³C chemical shifts.¹¹ For example, while structure 1a could arise from a [2 + 2] photocycloaddition of structure 3c in which the C2-C10 and C3-C9 bonds are formed, we considered structures that might arise from analogous cyclizations of the other asteriscunolide isomers or [2+2] reactions involving the C2=C3 and C6=C7 π bonds. In each case, we considered the bond-formation events that appeared most reasonable on the basis of the reported crystal structures of the asteriscunolide isomers.⁸ Such considerations produced structures ranging from reasonable to highly implausible on geometric grounds, but the computed chemical shifts for none of these were an acceptable match to the reported chemical shifts. Although we initially discounted the structure 1b (Figure 2) that might result from C2-C9/C3-C10 cyclization of 3c (the lowest-energy asteriscunolide isomer^{12,13}) because such a closure seemed unlikely on the basis of the crystallographic conformation of 3c, this structure could be formed from other conformations. To our delight, the computed chemical shifts for 1b matched the values reported for aquatolide

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| Tabl | le 1. (| Comparison of | Experimental | and Co | omputed NMR | Chemical | Shifts for A | Aquatolide |
|------|---------|---------------|--------------|--------|-------------|----------|--------------|------------|
| | | | | | | | | |

| | | ¹³ C NMR chemica | al shifts (ppm) | | ¹ H NMR chemical | shifts (ppm) | | |
|----------------------|-------------------------|-----------------------------|----------------------|--------------------|-----------------------------|----------------------|----------------------|--------------------|
| nucleus ^a | exptl ^b | comp 1a ^c | comp 1b ^c | exptl ^d | exptl ^b | comp 1a ^c | comp 1b ^c | exptl ^d |
| 1 | 84.12 | 80.76 | 83.29 | 84.20 | 4.48 | 4.23 | 4.21 | 4.48 |
| 2 | 54.48 | 41.66 | 54.75 | 54.54 | 3.25 | 2.99 | 3.20 | 3.26 |
| 3 | 62.77 | 48.84 | 62.83 | 62.83 | | | | |
| 4 | 22.07 | 31.23 | 22.46 | 22.15 | 2.50 | 1.99 | 2.48 | 2.52 |
| | | | | | 2.02 | 1.89 | 1.89 | 1.96 |
| 5 | 28.53 | 30.16 | 30.49 | 28.63 | 2.38 | 3.69 | 2.29 | 2.35 |
| | | | | | 2.07 | 2.32 | 2.08 | 2.03 |
| 6 | 130.94 | 143.61 | 135.22 | 131.10 | 5.84 | 6.36 | 5.94 | 5.85 |
| 7 | 135.07 | 139.04 | 138.11 | 135.08 | | | | |
| 8 | 211.58 | 211.93 | 211.87 | 211.94 | | | | |
| 9 | 54.37 | 55.53 | 54.76 | 54.45 | 2.92 | 3.22 | 2.78 | 2.92 |
| 10 | 62.57 | 38.24 | 63.89 | 62.59 | 2.64 | 2.97 | 2.49 | 2.64 |
| 11 | 41.73 | 44.89 | 44.57 | 41.86 | | | | |
| 12 | 177.27 | 181.17 | 177.65 | 177.50 | | | | |
| 13 | 22.00 | 21.98 | 23.19 | 22.22 | 1.86 | 1.89 | 1.84 | 1.87 |
| 14 | 22.73 | 16.72 | 21.09 | 22.62 | 1.18 | 0.99 | 0.98 | 1.05 |
| 15 | 22.46 | 25.21 | 20.80 | 22.84 | 1.04 | 1.17 | 1.09 | 1.19 |
| | CMAD ^e | 7.23 | 1.37 | | CMAD ^e | 0.35 | 0.10 | |
| | largest $\Delta \delta$ | 24.33 | 4.28 | | largest $\Delta \delta$ | 1.31 | 0.27 | |

^{*a*}For numbering, see Figures 1 and 2. ^{*b*}Data in CDCl₃ taken from ref 1. ^{*c*}Data computed for the originally proposed structure (**1a**) and the revised structure (**1b**); see the SI for details. ^{*d*}Present work. ^{*e*}CMAD is the corrected mean absolute deviation, computed as $(1/n)\sum_{i=1}^{n}|\delta_{\text{comp},i} - \delta_{\text{exptl},i}|$, where δ_{comp} refers to the scaled computed chemical shift.

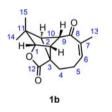


Figure 2. Revised structure of aquatolide.

exceptionally well (Table 1).¹⁴ Structures derived from the same type of cyclization for the other three asteriscunolide isomers did not produce an acceptable match to the experimental data.¹⁵

While structure **1b** does not possess a ladderane core, the ring system (bicyclo[2.1.1]hexane) is uncommon. Two notable examples that lack the lactone bridge are found in cyclo-pampeanone isovalerate (**4**)¹⁶ and solanoeclepin A (**5**),¹⁷ the latter of which has been subjected to X-ray crystallographic analysis (Figure 3). Although the biosynthesis of **5** is uncertain, the cyclopampeanone core is thought to arise from an oxidized bisabolene precursor that can undergo [2 + 2] cycloaddition or an intramolecular Alder ene reaction, the latter of which leads to the isomeric pampeanone isovalerate (not shown).¹⁶ All three metabolites are isolated from the same source, supporting the

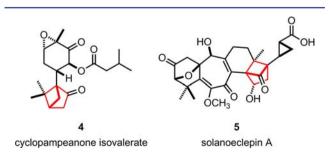


Figure 3. Known natural products with a bicyclo[2.1.1]hexane core.

proposed biosynthesis. The exact number of secondary metabolites that share the bicyclo[2.1.1]hexane core with aquatolide is difficult to discern. In view of the propensity of this core structure to exhibit unusual ${}^{1}\text{H}{-}^{1}\text{H}$ coupling in the NMR spectrum (see below), the number of structures is likely to grow as more are discovered or reassigned.

We next turned our attention to the experimental data reported by San Feliciano and co-workers.1 These authors proposed structure 1a largely on the basis of homonuclear and heteronuclear 2D NMR experiments. While it does appear that the proposed structure is fully consistent with the reported experimental data, the absence of a reported NOE correlation between H2 and H10 intrigued us, and we wondered whether additional NMR experiments might further support our proposed structural revision. We thus decided to attempt to reisolate the original compound and/or 3a-d, which could potentially serve as synthetic precursors to 1a or 1b. A. aquaticus was collected in March 2012 on the south coast of Mallorca, Spain, and then dried and shipped to UC Davis. The plant material (231.7 g) was ground and extracted with hexanes, and the crude hexanes-soluble extract (7.6 g) was chromatographed on silica gel. Fractions were analyzed by GC-MS for those containing m/z 246. Several of the asteriscunolides were isolated, as was a small quantity (2.5 mg) of a compound with a fragmentation pattern similar to that reported for aquatolide. The isolated compound showed an $[M + Na]^+$ ion at m/z269.1158 (calcd 269.1154) and the same ¹H and ¹³C NMR chemical shifts as reported by San Feliciano and co-workers¹ (Table 1). The observed melting point and optical rotation showed reasonable agreement with reported values as well [see the Supporting Information (SI) for details]. The IR spectrum showed strong bands at 1755 and 1674 cm⁻¹ consistent with the γ -lactone and α,β -unsaturated ketone, respectively. These data indicated that the compound isolated was the same compound that San Feliciano and co-workers isolated and named aquatolide.

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The ${}^{1}H-{}^{1}H$ coupling constants observed for the reisolated compound (Table 2) also matched those originally reported.

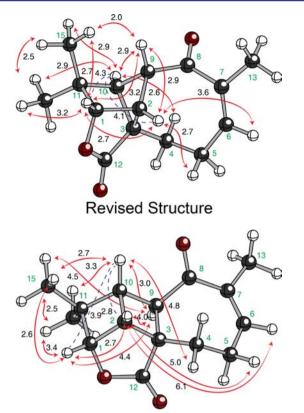
Table 2. Experimental and Computed ¹H-¹H Coupling Constants (in Hz) for Structure 1b

| nucleus ^a | $experimental^b$ | computed ^c |
|----------------------|---------------------|--|
| 1 | t (2.2) | $J_{\rm H2} = 2.3, J_{\rm H10} = 1.8$ |
| 2 | dd (7.3, 2.5) | $J_{\rm H1} = 2.3, J_{\rm H10} = 6.8$ |
| 4a | dd (15.8, 6.7) | $J_{4b} = 15.7, J_{5a} = 1.8, J_{5b} = 6.7$ |
| 4b | m | |
| 5a | m | |
| 5b | m | |
| 6 | ddt (4.7, 3.1, 1.5) | $J_{5a} = 3.4, J_{5b} = 5.4, J_{H13} = 1.8$ |
| 9 | s | $J_{\rm H2} = 0.1, J_{\rm H10} = 0.07$ |
| 10 | dd (7.3, 1.8) | $J_{\rm H1} = 1.8, J_{\rm H2} = 6.8, J_{\rm H9} = 0.07$ |
| 13 | q (2.0) | $J_{\rm H5a} = 2.5, J_{\rm H5b} = 2.8, J_{\rm H6} = 1.8$ |
| 14 | s | |
| 15 | S | |

^{*a*}See Figure 2. ^{*b*}Present work; these data are fully consistent with those reported in ref 1. ^{*c*}Selected *J* values only, reported as absolute values; see the SI for details and the complete set of computed values.

However, these data initially seemed to be at odds with structure 1b (and possibly a better fit for structure 1a). For example, H10 was found to be a doublet of doublets with J = 7.3 and 1.8 Hz. This hydrogen has two ³J neighbors in structure **1a** but only one in structure 1b. To understand this apparent discord, we computed the ¹H-¹H coupling constants for both structures using quantum-chemical techniques.⁶ The computed values for structure 1b (Table 2) displayed an excellent overall match to the experimental data (see the SI for the computed values for structure 1a, which did not match the experimental data). In fact, the computed results bring to light an atypically large ⁴*J* coupling of \sim 7 Hz between H2 and H10. While other ⁴J coupling constants in the molecule are not as large (e.g., $J_{\rm H1-H10} \approx 2$ Hz and $J_{\rm H1-H9} \approx 0$ Hz), all are consistent between experiment and theory in this case. Examples of comparably large ⁴J values in similar structures are known.¹⁸

We next examined data from several homonuclear and heteronuclear 2D NMR experiments. The ¹H-¹H COSY and $^{1}H-^{13}C$ HSQC spectra confirmed the connectivity among H1, H2, and H10 predicted by the computed ${}^{1}H-{}^{1}H$ coupling constants and previously reported for the original structure.¹ These data also confirmed the absence of coupling for H9, which is nearly orthogonal to both H2 and H10 (the computed dihedral angles are 80° and 85°, respectively). The HMBC spectrum of the central core of the molecule (H1, H2, H9, H10) was inconclusive. Although this experiment was optimized to show ³J heteronuclear coupling, the frameworks of both structures places these H atoms 2–4 bonds away from the relevant carbon atoms, making it difficult to draw any firm conclusions on the basis of these data. A NOESY experiment, on the other hand, was more conclusive. We observed the same NOE correlations as originally reported, and these appeared to be consistent with both possible structures.¹⁹ However, correlations between H9 and H10 and between H2 and H9, which are more consistent with structure 1b, were also observed. In particular, the observed H2/H9 correlation seems unlikely for structure 1a. In addition, an NOE correlation between H2 and H10 was not observed, a result that further supports the revised structure since the distance between these two atoms is computed to be >4 Å in structure 1b but <3 Å in structure 1a. Shown in Figure 4 are the observed NOE



Original Structure

Figure 4. Observed NOE correlations overlaid on the computed geometries (lowest-energy conformers) for the revised and original aquatolide structures. The value near each arrow is the distance between the two atoms in Å. Blue dashed lines are selected distances for which no NOE correlation was observed.

correlations from the present work overlaid on the original and revised structures of aquatolide. It is clear that the revised structure is more consistent with the observed correlations, with the longest NOE distance being 3.6 Å in the revised structure compared with 6.1 Å in the original structure.

The revised structure **1b** assigned on the basis of quantumchemical calculations and NMR experiments was subsequently confirmed by X-ray crystallography.²⁰ In addition to confirming the core connectivity, the crystallographic data confirmed the absolute configuration on the basis of the observed Flack parameter of -0.07(15).

The determination of the unusual core structure of aquatolide posed a significant challenge. Extensive NMR analysis, including a synergistic interplay between experiment and calculations, was required to elucidate the structure. Although the originally proposed structure was consistent with the set of data reported, it was not the only possible structure that could fit the data. In fact, certain features such as large four-bond ¹H-¹H coupling constants, small three-bond IH-1H coupling constants, and other unusual correlations found with this molecule initially seemed to support the original structure better. Such features could therefore easily lead researchers toward incorrect determinations in some cases.²¹ These pitfalls can be avoided by using quantum-chemical calculations of chemical shifts and coupling constants, especially in combination with the careful implementation of state-of-the-art NMR experiments. Such computational data were useful here not only for ruling out candidate structures and supporting the correct one but also for reconciling the unusual coupling constants and NOE data observed for aquatolide, further demonstrating the utility of quantum-chemical calculations in making structural assignments for small quantities of natural materials for which crystallography may be impossible or impractical.²²

ASSOCIATED CONTENT

S Supporting Information

Computational and experimental details. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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(11) In all, ~60 constitutional isomers were considered, and in most cases, various diastereomers of each were considered as well (see the SI).
(12) The bicyclo[2.1.1]hexane framework of 1b is expected to arise from a "crossed" nonconcerted cycloaddition from the T1 state (typical for enones). Crossed rather than parallel cycloaddition (the latter would lead to 1a) is expected to predominate for such systems. For example, see: (a) Srinivasan, R.; Carlough, K. H. J. Am. Chem. Soc. 1967, 89, 4932.
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(13) The computed gas-phase free energies of **3a**, **3b**, and **3d** relative to **3c** are 5.32, 7.30, and 2.11 kcal/mol, respectively (see the SI for details). (14) With corrected mean absolute deviations of 1.37 and 0.10 ppm for ¹³C and ¹H, respectively, the closeness of the match is consistent with applications of the same methods to other known complex natural product structures. See the SI for calculations on natural products of comparable complexity, including aplydactone, ingenane, intricarene, echinopine B, and naupliolide, as well as on bicyclo[2.1.1]hexan-2-one. Also see ref 5.

(15) (a) It thus seems that [2+2] photocycloaddition of 3c to form a bicyclo^[2,1,1] structural unit is the likely biosynthetic route to aquatolide. The alternative photocycloaddition to form the originally proposed [2]ladderane structure apparently does not occur to any appreciable extent, at least in A. aquaticus. Interestingly, $\begin{bmatrix} 2 + 2 \end{bmatrix}$ photocycloaddition of myrcene, which could also lead to structures with bicyclo[2.1.1]hexane or [2]ladderane cores, has been shown to produce only the bicyclo[2.1.1]hexane product (see ref 12b). (b) Notably, asteriscanolide may be related to the corrected structure of aquatolide as a product of its fragmentation or as a product of incomplete [2 + 2]reaction, photochemical or not (we thank Dr. Yong Liang of UCLA for sharing the results of his preliminary calculations on these possibilities). Also see: El Dahmy, S.; Jakupovic, J.; Bohlmann, F.; Sarg, T. M. *Tetrahedron* **1985**, *41*, 309. (c) The synthesis of a related bicyclo^[2.1.1] hexane system bearing a gem-dimethyl group and a fused lactone unit was described previously, but NMR data for this compound were not provided. See: Meinwald, J.; Gassman, P. G.; Hurst, J. J. J. Am. Chem. Soc. 1962, 84, 3722.

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(19) The only difference between the assignments for the original and revised structures involves the shifts for the methyl hydrogens H14 and H15, which were switched in the revised structure on the basis of the observed H9/H15 correlation. No correlation between H14 and H9 was observed.

(20) See the SI for full crystallographic details. The structural data for **1b** were deposited at the Cambridge Crystallographic Data Centre (CCDC) under deposition number 898298.

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